

# Growth of *Salmonella enterica* in model mixed cultures during a two-step enrichment

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## SUMMARY

Growth of *Salmonella enterica* was studied in model mixed cultures with *Citrobacter freundii* or *Escherichia coli* in buffered peptone water (BPW) and in Rappaport-Vassiliadis medium with soya (RVS) with modified concentrations of  $MgCl_2$  and malachite green, and at modified incubation temperatures. Selected *S. enterica* strains were inoculated in BPW ( $10^0$  cfu/ml) together with selected strains of *Citrobacter freundii* (up to  $10^8$  cfu/ml) or selected strains of *Escherichia coli* (up to  $10^8$  cfu/ml), incubated overnight and then subcultured (1: 100) in RVS variants. Growth of individual bacterial species was followed by the quantitative real-time polymerase chain reaction (PCR). Optimal culture conditions during the second selective step were:  $MgCl_2 \cdot 6 H_2O$  concentration of 29 g/l, malachite green concentration of 36 mg/l, and the incubation temperature of 41.5 °C. *Citr. freundii* was found to be a potent competitor and *E. coli* was a weaker competitor. At optimal culture conditions, competition was reduced and the density of *S. enterica* cultures reached the level of  $10^4$  cfu/ml after not later than 2 h of selective enrichment. The results obtained provide a basis for the development of a short two-step enrichment to be used in rapid real-time PCR-based methods for the detection of *S. enterica* in food and other matrices.

**KEY WORDS:** *Salmonella*, Rappaport-vassiliadis medium with soya (RVS), Real-time polymerase chain reaction (PCR), *Citrobacter*, *Escherichia coli*

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## INTRODUCTION

*Salmonella enterica* is a food-borne pathogen which represents a major public health problem in almost all industrialized countries (Tietjen *et al.*, 1995). For routine detection of *S. enterica* in food and other matrices, a conventional method according to EN ISO 6579 (2002) is used. This standard method is sensitive and reliable but requires up to 7 days. Recently, a number of rapid methods for the detection of *S. enterica* in food have been developed including immunoassays and polymerase chain reaction (PCR) tech-

niques. The best parameters were reported for methods based upon real-time PCR which detected the pathogen at the level of  $10^0$  cfu / 25 g in a closed-tube format (Uyttendaele *et al.*, 2003; Malorny *et al.*, 2004; Patel *et al.*, 2006).

Most currently available real-time PCR-based methods for the detection of *S. enterica* in food include an overnight pre-enrichment in buffered peptone water (BPW). This simple step seems to provide a sufficient amount of *S. enterica* for downstream detection (Malorny *et al.*, 2004). Nevertheless, the single-step enrichment may produce false positive results caused by the presence of dead bacterial cells in the sample (Drahovská *et al.*, 2001) or false negative results caused by food-borne PCR inhibitors (Salomonsson *et al.*, 2005). These problems may be solved if a two-step enrichment is used. In this case, dead bacterial cells as well as food-borne PCR inhibitors would be diluted and would not affect the results

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